

Contents lists available at ScienceDirect

Applied Soil Ecology



journal homepage: www.elsevier.com/locate/apsoil

Compensatory mechanisms of litter decomposition under alternating moisture regimes in tropical rice fields



Anja Schmidt^{a,*,1}, Katharina John^{b,1}, Harald Auge^{a,c}, Roland Brandl^d, Finbarr G. Horgan^{e,f}, Josef Settele^{a,c}, Andrey S. Zaitsev^b, Volkmar Wolters^b, Martin Schädler^{a,c}

^a Helmholtz Centre for Environmental Research – UFZ, Department of Community Ecology, Theodor-Lieser-Str. 4, 06120 Halle, Germany

^b Justus-Liebig-University Giessen, Department of Animal Ecology, Heinrich-Buff-Ring 26-32, 35392 Giessen, Germany

^c iDiv, German Centre for Integrative Biodiversity Research, Halle-Jena-Leipzig, Deutscher Platz 5e, 04103 Leipzig, Germany

^d Philipps-University Marburg, Faculty of Biology, Department of Ecology, Karl-von-Frisch-Str. 8, 35032 Marburg, Germany

^e International Rice Research Institute, DAPO 7777 Metro Manila, Philippines

^f Centre for Compassionate Conservation, University of Technology Sydney, 15 Broadway, Sydney NSW 2007, Australia

ARTICLE INFO

Article history: Received 14 January 2016 Received in revised form 13 May 2016 Accepted 19 May 2016 Available online xxx

Keywords: Litterbag Detritivore Oryza sativa The Philippines Soil fauna Crop rotation

ABSTRACT

A conversion from permanently flooded rice cropping systems to crop rotations that include non-flooded upland crops is heavily promoted in tropical regions to meet the challenges of sustainable food production while reducing water consumption and trace gas emissions. Shorter periods of flooding and manipulation of fertilizer inputs affect belowground community composition, biomass and functioning. However, there have been no previous studies of how such shifts in rice field management might affect soil biota and decomposition. Our objective was to examine how crop diversification, which demands different moisture regimes and nitrogen inputs, influences belowground invertebrate assemblages and their contribution to the decomposition of crop residues.

We conducted a litterbag experiment in lowland paddy fields at the experimental field sites of the DFG-ICON project (Laguna, the Philippines) that were either continuously flooded, had seasonally alternating wet and dry periods, or were continuously dry. Additionally, subplot treatments within these crop rotations included different N fertilizer management practices. At a total of 36 plots we used litterbags with two different mesh sizes to assess decomposition with and without fauna over a period of 72 days. Furthermore, we sampled soil microfauna and mesofauna in both the wet (rainy) and dry seasons.

Although we found no correlation between faunal abundance and the contribution by invertebrates to rice straw decomposition, we found that soil water content was the decisive factor determining the activity of decomposer invertebrates as well as the composition and abundance of the soil fauna in fields with alternating cropping regimes. The impact of invertebrates on rice straw decomposition was higher under anaerobic than aerobic soil conditions which compensated for reduced microbial decay rates during periods of flooding. In contrast, microbial decomposition rates were higher under aerobic conditions, whereas invertebrates had no apparent effects on the mass loss of rice straw despite their higher abundance in dry fields. Our results demonstrate that invertebrates are essential for the effective decay of rice straw residues under flooded soil conditions, and therefore play an important role in supplying nutrients to flooded rice.

© 2016 Elsevier B.V. All rights reserved.

1. Introduction

Flooded rice cultivation with two to three crops per year is the most predominant agricultural land use system in the lowland tropics and subtropics of Asia (Cassman and Pingali, 1995) and provides about 90 % of the global rice supply (FAOSTAT, 2008; USDA, 2007). Currently, the sustainability of producing rice in flooded fields has been questioned in regard to water use efficiency and greenhouse gas emissions (Bouman et al., 2007; Mueller et al., 2012). A range of management strategies have been developed to reduce water consumption and methane emissions from ricebased cropping systems while ensuring sufficient productivity (e.g. Wassmann and Vlek, 2004). One of the most promising approaches

* Corresponding author.

http://dx.doi.org/10.1016/j.apsoil.2016.05.014 0929-1393/© 2016 Elsevier B.V. All rights reserved.

E-mail address: a.schmidt@ufz.de (A. Schmidt).

¹ Authors contributed equally to the manuscript and share first authorship.

is the inclusion of non-flooded crops into rice-dominated systems (Bouman et al., 2005; Timsina et al., 2010, 2011). However, crop diversification in rice-based systems also bears the risk of depleting soil nutrients and reducing soil organic matter (Haefele et al., 2013; Witt et al., 2000).

A rich assemblage of organisms inhabits paddy rice soils (Bambaradeniya and Amarasinghe, 2003). These organisms significantly contribute to the decomposition of crop residues and element cycling (Lekha et al., 1989; Schmidt et al., 2015b; Widyastuti, 2002). The belowground community of conventionally managed paddy soils is dominated by aquatic and semi-aquatic oligochaetes and nematodes, whereas microarthropods are generally rare (Watanabe and Roger, 1985). Changes in water management by introducing upland crops together with consequent adjustments in fertilizer use are likely to adversely impact native assemblages of soil animals in rice fields. A reduced contribution by the soil fauna to decomposition processes could also diminish microbial activity and thus the mineralization of both carbon (Setälä et al., 1988) and nitrogen (Huhta et al., 1988). On the other hand, increased niche diversity over time due to crop diversification might stimulate the increase of abundance, richness and functional diversity of belowground fauna (Cancela da Fonseca and Sarkar, 1998) with positive implications for litter decomposition. For example, in continuously flooded fields soil mites and springtails have been shown to play a minor role in the decomposition of organic matter (Widyastuti, 2002), but under aerobic conditions their abundance rapidly increases (Widyastuti, 2002) which might enhance their contribution to the regulation of microflora-mediated decomposition.

Decomposition of soil organic matter is one of the most important ecological processes regulating nutrient cycling and energy flow in the soil (Lekha et al., 1989; Manzoni et al., 2008; Talbot and Treseder, 2011). It is highly interactive and involves a broad spectrum of micro-, meso- and macrofaunal groups (Anderson, 1995; Coleman and Crossley, 1996; Wolters, 1991) that generally have indirect effects on the decay of organic matter, e.g. by increasing the surface area and improving the quality of litter as a substrate for microbial colonization (Heath et al., 1964). Hence, microbial activity and biomass are strongly defined by higher trophic level interactions in soil food webs (Lekha et al., 1989; Singh et al., 2005). In addition, the rate of litter decomposition is triggered by several biotic and abiotic factors such as soil properties (e.g. soil moisture and soil texture; Amato and Ladd, 1992; Clark and Gilmour, 1983; Merckx et al., 1985; Steinberger and Whitford, 1988) and the biochemical composition of the plant residues (e.g. N availability and lignin concentration; Bollen, 1953; García-Palacios et al., 2013; Singh et al., 2005; Tian et al., 1993). Decomposer invertebrates are likely to increase the effectiveness of litter decomposition in flooded rice fields (Schmidt et al., 2015a) where microbial activity is hampered by the high C/N ratios and lignin contents of rice straw as well as the anaerobic conditions of flooded fields (Acharya, 1935; Koegel-Knabner et al., 2010; Schmidt et al., 2015b; Vigil and Kissel, 1991).

Different management practices can have a large impact on the organic C and N balance in the soils of continuous rice cropping and rice-maize rotation systems (Witt et al., 2000). Furthermore, the abundance and composition of soil fauna assemblages are influenced by crop type (Wardle et al., 1999). However, previous studies have not considered how shifts in soil conditions during crop rotations and consequent effects on the composition of soil animal assemblages might affect decomposition and soil nutrient dynamics. Recently, Handa et al. (2014) reported that a reduction in the functional diversity of decomposer organisms slows down the cycling of litter derived C and N across a range of terrestrial and aquatic ecosystems. Furthermore, an increase in plant species richness after the introduction of crop rotations affects the biomass

of microbes and soil animals (Wardle et al., 1999) which in turn might alter the rate of nutrient cycling in the soil. A better understanding of soil-dwelling animals and their impact on the dynamics of decomposition processes in rice-based agroecosystems will be essential for evaluating the benefits and risks associated with the introduction of non-flooded crops into these systems, and hence, for developing effective management strategies for the soils of rice producing regions.

The present study was conducted at the experimental field sites of the ICON (*"Introducing Non-Flooded Crops in Rice-Dominated Landscapes: Impact on Carbon, Nitrogen and Water Cycles"*) research group located in Los Baños (the Philippines). We examined how introducing non-flooded upland crops with different fertilization regimes into the cropping cycles of continuously flooded rice fields alters the assemblages of soil animals and the invertebrate-driven decomposition of rice straw. We addressed the following hypotheses:

Compared to non-flooded fields, microbial litter decomposition in flooded rice fields will decrease due to anaerobic soil conditions, which increases the relative importance of decomposer invertebrates.

The conversion from wet to dry conditions during crop rotation alters the structure of soil invertebrate assemblages. We expect macro- and microarthropod groups to be more abundant under aerobic field conditions.

The lack of aquatic invertebrates and their contribution to decomposition processes in soils of non-flooded fields is compensated by a more abundant and active terrestrial mesofauna.

2. Materials and methods

2.1. Study site

Experiments were conducted at the International Rice Research Institute (IRRI) about 66 km south of Manila (Laguna, the Philippines; elevation: approx. 20 m above sea level; coordinates: lat 14.2, lon 121.4 WGS84 decimal degrees). In this region lowland flooded rice is primarily cultivated in two crop cycles per year, one in the dry season (December–May) and one in the wet season (June–November). Our study was carried out during both seasons in 2013. The site had been used for paddy rice cultivation for several decades prior to the establishment of the field experiment by the ICON group.

Average minimum, mean and maximum temperatures during the time of our experiment (2012/12–2013/11) were 24.0, 27.7 and 31.5 °C. The climate is characterized by a high intra-annual variation of precipitation. The average rainfall per month during the dry season (2012/12–2013/05) was 87.4 mm and average monthly rainfall during the wet season (2013/06–2013/11) was 343.0 mm.

The soil is of volcanic origin and clay-dominated. It is classified as Andaqueptic Haplaquoll (USDA classification) with 54.2 % clay, 32.8 % silt and 13 % sand. The bulk density of the topsoil (0-0.1 m) is 1.0 kg dm⁻³, the average pH is 6.1 and the organic C content is about 1.8 %.

2.2. Study design

Fieldwork was conducted at the experimental site (36 plots; 4 crop rotations \times 3 fertilizer treatments \times 3 replicates) of the ICON project ("Introducing Non-Flooded Crops in Rice-Dominated Landscapes: Impact on Carbon, Nitrogen and Water Cycles"). The 12 core fields (530–549 m² each, total area of about 150 \times 100 m) are arranged in two rows with six fields in each row (see Fig. 1 for the core field design). The fields were separated by field bunds and encompassed by grass verges (5 m in diameter). Since the dry



Fig 1. Setup of the ICON field experiment ("Introducing Non-Flooded Crops in Rice-Dominated Landscapes: Impact on Carbon, Nitrogen and Water Cycles"; Los Baños, the Philippines; scheme of the core field design). The 12 fields ($530-549 \text{ m}^2$ each) are arranged in two rows. The fields are managed with four different crop rotations (R-WET: flooded rice in dry and wet season; R-MIX: aerobic rice in dry season – flooded rice in wet season; M-MIX: maize in dry season – flooded rice in wet season; M-DRY: maize in dry season – aerobic rice in wet season). Each field is split into two plots ($159-197 \text{ m}^2$ each) with three different N fertilizer management regimes (z: no N fertilization; c: conventional N fertilizer application (130 kg N ha^{-1}); s: sitespecific N fertilizer application (180 kg N ha^{-1})).

season 2012, the fields had been managed with four different cropping systems. One was continuously flooded to represent the traditional double-rice cropping system with two seasons of lowland flooded rice per year (*Oryza sativa* L., NSIC Rc222, Tubigan 18; henceforth abbreviated by 'R-WET', see Table 1). In two intermittently submerged rotations, flooded rice during the wet season was followed by an upland crop during the dry season which consisted of either aerobic rice (NSIC Rc192, Sahod Ulan 1; 'R-MIX') or maize (*Zea mays* L., Pioneer hybrid P3482YR; 'M-MIX'). The fourth treatment was a continuously aerobic crop rotation where maize was grown in the dry season and aerobic rice in the wet season ('M-DRY').

Each field was split into three plots; each with a similar surface area (159–197 m²) separated by field bunds, and managed with three different N fertilizer treatments which were randomized over two parallel rectangles of about 24×6 m and one square plot of approximately 12×12 m. One plot per field received no N fertilization ('zero N'), the second one was conventionally fertilized

Table 1

Crop rotations of the ICON field experiment ("Introducing Non-Flooded Crops in Rice-Dominated Landscapes: Impact on Carbon, Nitrogen and Water Cycles"; Los Baños, the Philippines) and the respective crops cultivated in the dry and the wet seasons.

	Crop rotation	
Acronym	Dry season	Wet season
R-WET	Flooded rice	Flooded rice
R-MIX	Aerobic rice	Flooded rice
M-MIX	Maize	Flooded rice
M-DRY	Maize	Aerobic rice

('conv N', 130 kg N ha⁻¹) based on local farmers' practices, and the third treatment was fertilized following the site-specific N fertilization method ('site-spec N') which represents a dynamically adjusted nutrient management strategy where fertilizer inputs are calculated with the IRRI 'Rice Crop Manager' (http://webapps.irri. org/ph/rcm/) for site-specific conditions (estimated as 180 kg N ha⁻¹). Critical time points for fertilizer application were determined with the IRRI leaf color chart according to the chlorophyll content (LCC, Witt et al., 2005). N fertilizer was applied as urea three times a season. Additionally, muriate of potash and Solophos were applied at seasonal rates of 30 kg ha⁻¹ K₂O and 30 kg ha⁻¹ P₂O₅ for maize, respectively.

Rice seedlings (flooded and aerobic rice) were sown to seed beds in a greenhouse and transplanted to the fields 21 days after seeding (2–3 seedlings per hill with 0.2×0.2 m spacing). Flooded rice fields were kept submerged (approx. 0.05 m water body) after crop establishment (31 days after seeding) until two weeks before harvest. Land preparation consisted of soaking, plowing and puddling as well as harrowing and leveling. Maize and aerobic rice were grown in non-puddled soil. Fields were irrigated when rainfall was insufficient. Drainage ditches prevented fields with upland crops from flooding during heavy rains. Maize was planted with 0.25×0.75 m seed spacing.

After harvest maize plants were cut to ground level and removed from the fields with root residues remaining in the field (110 days after sowing). Rice plants were cut to approx. 5 cm above the soil level (aerobic rice: 89 days after transplanting; flooded rice: 100 days after transplanting). All residues were incorporated into the soil during land preparation for the next plant growth season. Manual weeding and rodent control were performed once per season as well as molluscicide (organic "Kuhol buster" – saponin) and fungicide ("Score") application.

2.2.1. Litterbags

In order to quantify the contribution by decomposer invertebrates to total rates of decomposition, nylon litterbags $(15 \times 20 \text{ cm})$ with two different mesh sizes (Bokhorst and Wardle, 2013) were filled with 10 g of litter (air-dried, chopped rice straw; Oryza sativa L., variety NSIC Rc222) and subsamples were retained for initial moisture and chemical analyses. The fine-meshed litterbags had a mesh size of $20 \times 20 \,\mu\text{m}$ and allowed access of microbes and some microfauna (protozoans, micro-nematodes) while the coarsemeshed bags had a mesh size of 5×5 mm. Tian et al. (1992) found no difference in rates of rice straw decomposition between mesh sizes of 2×2 mm and 7×7 mm. Henceforth, we will refer to the litter mass loss in fine-meshed bags as 'microbial decomposition'. Litter mass loss from coarse-meshed litterbags will be ascribed to the combined effects of microbial decomposition and faunal contribution to straw breakdown. Following Bradford et al. (2002) we consider the physical breakdown of plant material by the decomposer fauna and the consequent loss of small straw fragments from coarse-meshed bags as a functional role of decomposer invertebrates in the decomposition process and as part of the catabolic degradation process (Schädler and Brandl, 2005; but also see Frouz et al., 2015). Two weeks after transplanting or seeding (dry season: 2013/01/23; wet season: 2013/07/ 09) one fine- and one coarse-meshed litterbag were embedded in the upper soil layer in the middle of each field (5-10 cm below the soil surface) with a minimum spacing of 20 cm between bags (total number of bags: 72 per season). Litterbags were retrieved after 72 days of exposure in the field (approx. two weeks before harvest; DS: 2013/04/05; WS: 2013/09/19). After retrieval of the bags, soil particles, roots, and other alien plant material adhering to the straw were removed. The cleaned straw was dried at 60 °C for at least three days and weighed to the nearest centigram to calculate litter mass losses. The C and N contents of the original straw as well as retrieved straw from each litterbag were determined using an 'Elementar Vario EL element analyzer' (Elementar Analysengeräte GmbH, Hanau, Germany).

2.2.2. Soil sampling

Soil biota was sampled from all 36 plots. The field sampling was carried out twice each season (DS: 2013/01: WS: 2013/07: in total 144 sample sets). On each date, five soil core subsamples (approx. Ø 2 cm, 10 cm depth) were taken per plot for the extraction of nematodes following a modified Cobb's decanting and sieving method (Cobb, 1918). The nematodes were identified to genus level and assigned to feeding groups (Yeates et al., 1993). For the mesofauna, two soil cores (approx. Ø 5 cm, 10 cm depth) were taken. Following the extraction by means of the MacFadyen highgradient method (MacFadyen, 1961) microarthropods were sorted, counted and identified to suborder or family level. The second soil core sample was manually sieved and decanted for the extraction of Enchytraeidae (potworms) which were then suspended in 70 % ethanol and counted. Additional cores (Ø 5 cm, 10 cm depth) were used for the analyses of soil water content and total organic C content by the dry combustion method. As a proxy for fungal infestation, ergosterol, a fungal cell-membrane component, was measured according to Djajakirana et al. (1996). It was extracted from field-moist soil of 1 g dry weight with 100 ml distilled ethanol. The solution was shaken at $250 \text{ rev} \text{min}^{-1}$ for 30 min. Ergosterol was quantified by reversed-phase HPLC with 100 % methanol as mobile phase and detected at a wavelength of 282 nm (Heinze et al., 2010).

2.3. Data analysis

We carried out split-split plot ANOVAs using general linear mixed models (GLMM) Type III sum of squares (procedure MIXED, SAS 9.2) to analyze litter-, C- and N mass loss as well as the relative contents of C and N in dependence on crop (crop rotation; 4 levels within main plot), fert (fertilizer treatment; 3 levels within sub plot) and mesh (mesh size of litterbags; 2 levels within sub-sub plot) as well as their interactions. The factors field(crop) (main plot error) and $fert \times field(crop)$ (sub plot error) were introduced as random factors. Dry and wet season were analyzed separately to account for the fundamentally different climatic conditions and to allow independent tests of the effect of the current status of flooding in the different crop rotations on the observed variables. C and N mass loss were calculated by including the straw dry weight and its relative C and N contents, respectively, before setting and after retrieving of litterbags. To describe the sole influence of decomposer invertebrates on litter mass loss (by excluding the proportion of microbial decomposition on litter mass loss in coarse-meshed bags) the log response ratio was calculated (LRR; ln of litter mass loss in coarse-meshed bags divided by litter mass loss in fine-meshed bags).

Using the same statistical procedure, soil fauna data and ergosterol content were analyzed (split–split plot ANOVA) according to *crop* (crop rotation; 4 levels within main plot), *fert* (fertilizer treatment; 3 levels within sub plot) and *time* (soil core sampling date; 4 levels within sub–sub plot for soil fauna/2 levels within sub–sub plot for ergosterol) as well as their interactions. Again the terms *field(crop)* (main plot error) and *fert* × *field(crop)* (sub plot error) were introduced as random factors. Contrasts and post-hoc Tukey's HSD tests were carried out to reveal significant differences between the respective factor levels within factors.

To analyze community structures soil mesofauna and nematode abundances were calculated as individuals per m² and standardized using z-transformation. To reveal relations between assemblages of soil fauna abundance and the crop rotations (*crop*; categorical, 4 levels) within the two seasons (*time*; categorical, 4 levels) a redundancy analysis – RDA (Rao, 1964; van den Wollenberg, 1977) was carried out using R 2.1.4.2., package *vegan* (Oksanen et al., 2007). The variable *fert* (categorical, 3 levels) did not significantly influence the community structure and was therefore removed from the model. We used this specific multivariate method, which requires linear relationships between faunal groups as well as between assemblages and environmental variables (Euclidean metric), because of the homogeneity in our dataset and the short environmental gradient (Dormann and Kühn, 2008; Leyer and Wesche, 2007). According to Lepš and Šmilauer (2003) the use of linear methods is appropriate, if the longest gradient calculated using DCA/DCCA is smaller than 3. In our dataset the longest gradient was 1.9.

Analyses of co-variance (ANCOVA) were used to analyze the relationship between litter mass losses and selected soil-dwelling fauna groups including the same fixed and random factors as in the above described ANOVAs on litter mass losses. Faunal groups were included successively as covariates to reveal linear relationships of variances.

Structural equation models (SEM) are generally used to test for direct and indirect interaction effects between independent and measured variables in a single model (Grace, 2006). Seven observed endogenous variables were included in the model (SPSS Amos 22.0.0): soil water content, ergosterol content, log response ratio (LRR) of litter mass loss (as a proxy for invertebrate driven decomposition rates) and the abundance of Oribatida, Collembola, bacterial feeding Nematoda and fungal feeding Nematoda, respectively. Two categorical exogenous predictor variables (crop rotation and fertilizer treatment) with four and three factor levels. respectively, were transformed into dummy variables to include them in the model. To achieve this, a categorical variable with kdistinct values has to be converted into (k-1) dummy variables with binary coding [0,1]. Therefore, crop was transformed into three variables (M-DRY, M-MIX, R-MIX) and fert into two variables ('conv' - conventional fertilization, 'site' - site specific fertilization). Dummy coding of categorical variables with more than two levels compares the respective variables with the remaining "uncoded" control variable. Thus, the effect of one dummy variable on an endogenous response variable has to be interpreted in relation to the one variable not included in the model (R-WET for crop and 'zero' N for *fert*). For example, if a dummy variable has a negative regression weight of -0.4 on a response variable, the effect of the dummy variable is 40 % more negative than the effect of the control variable. As dummy variables are not independent of each other they need to be co-varied in the model. Model fitting was performed by the stepwise removal of insignificant and weak relationships with confirmation of adequate model fits by a decrease of CAIC (Bozdogan, 1987) of more than 2 (Grace, 2006) and non-significant χ^2 -tests (*P*>0.05). However, in the wet season we observed only low abundances of the decomposer taxa and therefore excluded the SEM for this season.

3. Results

3.1. Litter decomposition

The mass loss of rice straw from coarse- and fine-meshed litterbags was affected by the four crop rotations (significant $crop \times mesh$ interactions, Table 2). In both seasons, flooded rice fields had higher litter mass loss and higher N content in the straw retrieved from coarse-meshed bags compared to the fine-meshed ones (Fig. 2 A–D). Independent of the respective crop rotation, no significant impact of mesh size on these two parameters could be established during the aerobic stage (Fig. 2A–D). However, compared to the initial conditions, the relative N content generally

Table 2

The effects of *crop, fert, mesh* and their interactions on litter mass loss of rice straw and the N and C contents of the retrieved straw using a GLMM type III sum of squares. Significant effects are indicated in bold font.

	Litter mass loss (%)			N content	t (%)		C content (%)		
Factors	Df	F	Р	Df	F	Р	Df	F	Р
Dry Season									
crop	3,8	1.89	0.21	3,8	8.31	0.01	3,8	12.7	0.002
fert	2,16	0.25	0.78	2,16	4.67	0.03	2,16	0.16	0.86
mesh	1,24	9.54	0.005	1,24	5.60	0.03	1,24	38.8	<0.0001
crop imes fert	6,16	0.67	0.68	6,16	1.49	0.24	6,16	0.29	0.93
crop imes mesh	3,24			3,24	23.1	<0.0001	3,24	10.3	0.0001
fert \times mesh	2,24	0.46	0.64	2,24	0.36	0.7	2,24	2.45	0.11
crop imes fert imes mesh	6,24	0.14	0.99	6,24	1.80	0.14	6,24	0.90	0.51
Wet Season									
crop	3,8	2.96	0.1	3,8	4.64	0.04	3,8	0.80	0.53
fert	2,16	1.16	0.34	2,16	0.17	0.84	2,16	0.91	0.42
mesh	1,22	390	<0.0001	1,22	213	<0.0001	1,22	5.58	0.03
crop imes fert	6,16	0.25	0.95	6,16	0.61	0.72	6,16	0.63	0.7
crop imes mesh	3,22	28.1	<0.0001	3,22	10.9	0.0001	3,22	7.20	0.001
fert $ imes$ mesh	2,22	0.20	0.82	2,22	0.58	0.57	2,22	0.92	0.41
$crop \times fert \times mesh$	6,22	0.48	0.82	6,22	0.76	0.61	6,22	0.43	0.85

Factor *crop* represents the 4 different crop rotations (R-WET, R-MIX, M-DRY), the factor *fert* is the effect of the three different fertilizer treatments (zero, site specific, conventional), and factor *mesh* the two mesh sizes (5 mm and 20 μ m) used in every plot. The model also includes the random effects of *field(crop)* (crop nested in field) and *fert* × *field(crop)*; their effects and interactions are not shown.

increased during the decomposition process (Fig. 2 C+D; initial N content of the straw = 0.6 %). Contrast tests of the $crop \times mesh$ interaction comparing the state of flooding in the particular season (flooded vs. non-flooded; P < 0.05) revealed that the flooding regime had a much stronger impact on litter mass loss and N content (for both mesh sizes) than the mode of crop rotation. In contrast, C content was significantly different in non-flooded fields and identical under flooded conditions (Fig. 2 E+F). However, the decomposition process did not consistently alter the C content (Fig. 2 E + F; initial C content of the straw = 36.8 %). Total losses of C and N were smallest from fine-meshed bags placed in rice fields that were flooded in that particular season (Table A.1; Fig. A.1 A-D in Supplementary materials). The highest amounts of C and N were lost from coarse-meshed litterbags of flooded fields as well as from litterbags of both mesh sizes of non-flooded fields. No significant difference in the range of depletion of these elements was evident (Fig. A.1 A–D in Supplementary materials). C/N ratios in litterbags with different mesh size and placed in different crop rotations mirror this pattern with lowest values in straw residues of coarsemeshed bags in all fields as well as of fine-meshed bags during dry cropping. Highest values were found in fine-meshed bags during flooded conditions (Table A.1; Fig. A.1 E+F in Supplementary materials). With the single exception of the N content in the dry season, no significant effects of the factor fert or its interaction with the factors crop and mesh on mass losses of rice straw, C and N as well as on C and N content were detected (Table 2 and Table A.1 in Supplementary materials).

Ergosterol content in the soil indicates a negative correlation between fungal infestation and litter mass loss in fine-meshed bags ($r^2 = 0.25$; $P \le 0.001$; Fig. 3A). Moreover, fungal infestation was positively correlated to the log response ratio of litter mass loss ($r^2 = 0.29$; $P \le 0.001$; Fig. 3B; ANOVA table see Table A.2 in Supplementary materials).

3.2. Invertebrate fauna

Mites and collembolans were the most abundant groups, with the highest abundance recorded in the dry season (Fig. 4A + B). The factors *crop* and *time* as well as their interaction significantly affected the abundance of all selected soil mesofaunal groups (except for *crop* on springtails; Table 3). The mode of crop rotation significantly altered the abundance of Acari in the way that the highest values were recorded in M-DRY and the lowest in R-WET at the end of the dry season (Fig. 4A). Crop rotations also affected the abundance of springtails (Fig. 4B). However, this effect was confined to the beginning of the dry season, with the highest values occurring in M-MIX and the lowest in R-WET. In general, the abundance of the soil mesofauna was very low in R-WET and at all fields during the wet season (Fig. 4C). Fertilization significantly affected soil mesofauna abundance only in interaction with time (Table 3).

According to the redundancy analysis, the first RDA axis explained 8 % (Table A.3 in Supplementary materials; P=0.005) of the total variance and mostly represented the 'DS-end' (end of dry season) level of the variable time (Fig. 5; see also Table A.4 in Supplementary materials - highest absolute value at RDA 1). The second axis accounted for 4 % (Table A.3 in Supplementary materials; P = 0.005) of variance and was related to the level 'DSstart' (start of dry season) of the variable time (Fig. 5; see also Table A.4 in Supplementary materials – highest absolute value at RDA 2). In total, 14 % of the variance in the dataset was explained by the six constrained RDA axes. Of this variance, 57 % was explained by RDA 1 and 26 % by RDA 2 (Table A.3 in Supplementary materials). The two environmental factors crop and time significantly influenced the abundance of different soil invertebrate groups (both *P*=0.005; all results of ANOVA permutation tests are given in Table A.5 in Supplementary materials). Analyses of covariance revealed no significant relationship between litter mass loss from litterbags and the abundance of different soil invertebrate groups.

Fitting of the structural equation model (SEM) required the stepwise removal of unimportant relationships (regression lines) until the best fitted model was achieved (model fit indices of fitted model: χ^2 = 11.0, Df = 28, *P* = 0.998, CAIC = 134.75; for initial model see Fig. A.2 in Supplementary materials). The fitted SEM (Fig. 6; for regression weights see Table A.6 in Supplementary materials) revealed that 70 % of the variation in litter mass loss caused by invertebrates can be explained by the net effects of its predictors. The strongest impact on litter mass loss was mediated through soil water content (Table A.6 in Supplementary materials - highly significant positive regression weight of 0.72) which in turn was solely determined by the three dummy variables describing the crop rotations (M-DRY, M-MIX, R-MIX; all three rotations with dry cropping during the dry season). All three path coefficients had negative correlation values, hence R-WET most positively affected soil water content (see Fig. A.3 in Supplementary materials for



Fig. 2. Percent litter mass loss (A/B), N content (C/D) and C content (E/F) (means + standard error SE) of rice straw retrieved from fields with different crop rotations (*crop*, for abbreviations see Table 1) in coarse-meshed (decomposition by invertebrates and microorganisms) and fine-meshed (decomposition by microorganisms) litterbags. A, C, E: dry season; B, D, F: wet season. Hatched bars represent the aerobic variants in the particular season. Different letters above the bars indicate significant differences between means (Tukey's HSD, $P \le 0.05$). Values of the original straw: N = 0.6 %, C = 36.8 %.

details; ANOVA results are summarized in Table A.7 in Supplementary materials). No direct or indirect effect of fauna abundance on invertebrate-driven litter decomposition in the dry season could be detected. The abundance of bacterial feeding nematodes was reduced by the M-DRY treatment with a path coefficient of -0.45, while the abundance of Oribatida was supported by M-DRY (regression weight: 0.51). This corresponds to the ANOVA results (Fig. 4). Compared to the zero N management, conventional and site specific fertilization treatments significantly increased the

abundance of bacterial feeding nematodes with path coefficients of 0.35 and 0.41, respectively. With a regression weight of -0.44, soil water content had a direct negative effect on the total abundance of collembolans, while conventional fertilization of fields (compared to zero N) significantly increased the abundance of this taxon (regression weight: 0.32).

Analyses of co-variance with LRR of litter mass loss as independent variable and different fauna groups as co-variables (abundance of nematodes – bacterial feeding, fungal feeding,



Fig. 3. Linear regressions between (A) microbial contribution to litter mass loss (fine-meshed bags) with the ergosterol content in the soil (μ g per g soil dry mass); and (B) the log response ratio of litter mass loss and ergosterol content in the soil. Both graphs present data collected in the dry and the wet season 2014 from the ICON ("Introducing Non-Flooded Crops in Rice-Dominated Landscapes: Impact on Carbon, Nitrogen and Water Cycles") experimental plots representing different crop rotations with cultivation of either flooded rice, aerobic rice or maize (for details on crop rotations and the experimental setup see Table 1 and Fig. 1); $P \le 0.001^{***}$.

plant-feeding and omnivorous nematodes; mesofauna – Collembola, Acari) showed no direct relation between the measured litter mass loss assigned to decomposer invertebrates and their abundance in either season. Even though the abundance of the analyzed soil fauna taxa was lowest in irrigated rice fields and abundance data does not promote direct evidence for invertebrate activity increase, the relative change of C and N content in the straw of litterbags incubated in the soil indicate the contribution of invertebrates to rice straw decomposition during irrigated rice cultivation.



Fig. 4. Abundance of soil mites (A) and springtails (B) per crop rotation (*crop*, for abbreviations see Table 1) at the four sampling times; panel (C) shows the total number of all mesofaunal groups (number of individuals, means +SE). Hatched bars represent the aerobic variants in the particular season. Different letters above the bars indicate significant differences between means (Tukey's HSD, $P \le 0.05$). Post-hoc comparisons were only made within one point in time, not between sampling times.

Table 3

Factors	Acari/m ²			Collembola/m ²			Abundance of all mesofaunal groups/m ²		
	Df	F	Р	Df	F	Р	Df	F	Р
crop	3,8	8.44	0.01	3,8	2.43	0.14	3,8	9.26	0.01
fert	2,16	2.63	0.1	2,16	0.95	0.41	2,16	3.21	0.07
time	3,72	53.9	<0.0001	3,72	16.1	<0.0001	3,72	56.7	<0.0001
$crop \times fert$	6,16	0.97	0.48	6,16	2.71	0.05	6,16	1.48	0.25
crop × time	9,72	5.56	<0.0001	9,72	3.45	0.001	9,72	4.29	0.0002
fert × time	6,72	2.25	0.04	6,72	2.40	0.04	6,72	2.84	0.02
crop \times fert \times time	18,72	0.58	0.9	18,72	1.52	0.11	18,72	0.65	0.85

The effects of *crop, fert, time* and their interactions on selected mesofaunal groups using a GLMM type III sum of squares. Significant effects are indicated in bold font.

Factor *crop* represents the 4 different crop rotations (R-WET, R-MIX, M-DRY), the factor *fert* is the effect of the three different fertilizer treatments (zero, site specific, conventional), and factor *time* the effect of the four different points in time the mesofauna was collected (DS-start, DS-end, WS-start, WS-end). The model also includes the random effects of *field(crop)* (crop nested in field) and *fert* × *field(crop)*; their effects and interactions are not shown.

4. Discussion

Our findings close a gap in scientific knowledge on the role of invertebrates in the decay of crop residues and on the processes of fauna-mediated carbon and nitrogen turnover in tropical paddy fields (but see e.g. John et al., 2015; Schmidt et al., 2015a; Settle et al., 1996). The results of the present study confirm that

decomposer invertebrates are key organisms for promoting the effective decay of organic matter in rice-based agroecosystems. This was mainly indicated by the much higher mass loss of rice straw in coarse-meshed litterbags compared to fine-meshed bags, which corroborates the results of earlier field studies conducted under similar conditions (e.g. Schmidt et al., 2015a,b; Widyastuti, 2002) and supports our first hypothesis that invertebrates



Fig. 5. Euclidean distance biplot based on a redundancy analysis (RDA); faunal groups of soil samples are represented by their 4- letter abbreviations (see below). Site scores are shown with different shapes depending on their affiliation to the particular crop rotation (*crop*, for abbreviations see **Table 1**), and different colors illustrate the corresponding point in time of each season (*time*). Treatment arrows represent the factor *time*. Axis 1 explains proportionally 8 % ($P \le 0.01$) of the variation in the dataset; Axis 2 accounts for 4 % ($P \le 0.01$) of the variation. **Abbreviations of faunal groups**: Ench – Enchytraeidae; *Acari*: Orib – Oribatida, Gama – Gamasina, Acar – remaining Acari from soil samples (others than Orib and Gama); *Collembola*: Smin – Sminthuridae, Isot – Isotomidae, Podu – Poduromorpha, Ento – Entomobryomorpha; *Bacterial feeding nematodes*: Acro – *Acrobeles* spp., Acoi – *Acrobeloides* spp., Ceph – *Cephalobus* spp., Euce – *Eucephalobus* spp., Pana – *Panagrolaimus* spp., Plec – *Plectus* spp., Rhab – *Rhabditis* spp.; *Fungal feeding nematodes*: Apoi – *Aphelenchoides* spp., Aphe – *Aphelenchus* spp., Neot – *Neotylenchus* spp., Hirs – *Hirshmanniella* spp., Long – *Longidorus* spp., Euce – *Pratylenchus* spp., Hirs – *Hirshmanniella* spp., Long – *Longidorus* spp., Prat – *Pratylenchus* spp., Roty – *Rotylenchus* spp., Tyle – *Tylenchus* spp. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)



Fig. 6. SEM (Structural Equation Model; see Materials and methods 2.3). The fitted model presents data collected in the dry season from the ICON ("Introducing Non-Flooded Crops in Rice-Dominated Landscapes: Impact on Carbon, Nitrogen and Water Cycles") experimental plots representing different crop rotations with cultivation of either flooded rice, aerobic rice or maize (for details on crop rotations and the experimental setup see Table 1 and Figure 1); model fit indices: $\chi^2 = 11$, Df = 28, P = 0.99, CAIC = 134.75; numbers above factors and regression lines represent standardized estimates of explained variance and regression weights; higher regression weights are indicated by thicker lines, for details on regression weights see Table A.6 in Supplementary materials; red lines represent negative path coefficients, green lines represent positive paths, dashed lines represent non-significant paths; "Nematoda – bact" = bacterial feeding nematodes; "Collembola" = total abundance of all groups of collembolans; M-DRY, M-MIX, R-MIX = crop rotations (see Table 1); "Conv" = conventional fertilization treatment; "Site" = site specific fertilization treatment; "LRR litter mass loss" = log response ratio of litter mass loss. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

contribute to the decomposition of rice straw in paddy fields. Although the abundance of soil fauna was low in flooded rice fields, evidence for invertebrate activity in decomposition processes was provided by the relative change of C and N content in the straw that had been retrieved from the litterbags. Nitrogen concentration in straw retrieved from fine-meshed bags of flooded fields did not differ from the initial reference values. Similarly, there was no reduction in carbon concentration in straw from either type of litterbag. In contrast, the increase of nitrogen concentration in rice straw retrieved from coarse-meshed litterbags accounts for a comparably faster release of other components, such as silicon and potassium (Christensen, 1985; Saha et al., 2009). Nevertheless, increased nitrogen concentration along with constant relative carbon content also points to a faster loss of C compared to N. As this pattern can only be found in straw retrieved from coarsemeshed bags of flooded fields, it can be ascribed to the activity of the decomposer invertebrates. Hence, under flooded conditions the loss of carbon is probably triggered by the soil fauna, whereas the breakdown of nitrogen seems to be primarily microbial-driven. Furthermore, the higher absolute release of nitrogen from straw in coarse-meshed litterbags compared to fine-meshed ones suggests that invertebrates had a stimulatory effect on the activity of microdecomposers. Microbial decay rates are highly influenced by the availability of C and N in litter and soil (Jansson, 1958; Knapp et al., 1983; Tian et al., 1995). Relatively high C/N ratios in the residues of rice straw (approx. 61:1, see S 2) compared to other herbaceous plants (ranging from 19:1 to 30:1; see e.g. Schädler et al., 2003) may hamper the efficiency of microorganisms during decomposition. However, by breaking down the plant material into smaller pieces and mixing these with soil, decomposer invertebrates increase the residue-soil area of contact, creating a more stable and favorable environment for microbial colonization (Cogle et al., 1987; Singh et al., 2010). In turn, the microbiota colonizing straw increase its nutritional value (Graca, 2001; Martinez et al., 2014; Perez et al., 2014) and thus create a more attractive resource for detritivores (Bradford et al., 2002). This is supported by our results showing the positive correlation between fungal biomass (measured as ergosterol content) and invertebrate-driven rice straw decomposition (log response ratio of litter mass loss) under flooded conditions. In general, microbial mineralization processes are known to be limited in anaerobic environments like in flooded paddy soils (Acharya, 1935); however, invertebrates completely compensated for reduced microbial activity under these conditions.

Under aerobic conditions, microbial decomposition rates were relatively constant during the whole year as indicated by similar levels of mass loss from litterbags placed in dry fields during both seasons. However, fungal biomass was considerably higher in the wet season. Hence, microbial decomposition during dry cropping might have been primarily driven by bacteria, whereas fungi play an apparently greater role in the decay processes during flooded phases. This is in agreement with previous studies which identified fungi as major drivers of microbial decomposition under anaerobic conditions in soil (Baldy et al., 2007; Bergfur and Friberg, 2012). Reduced microbial decomposition under the unfavorable conditions in flooded paddy fields reaffirms the key role of decomposer invertebrates in crop residue decay through both direct and indirect effects (Reddy et al., 1994; Santos and Whitford, 1981; Widyastuti, 2002).

A number of studies across a range of agroecosystems have demonstrated the impact of soil fauna on litter decomposition (e.g. Gießelmann et al., 2010; Schädler and Brandl, 2005; Wall et al., 2008). However, the results of similar research in other cropping systems cannot be simply extrapolated to rice paddies as regular flooding creates a unique habitat that harbors quite peculiar assemblages of decomposers in both the water and the top soil (Bambaradeniya and Amarasinghe, 2003). Decomposer invertebrates in flooded rice fields are mostly represented by (semi-) aquatic and edaphic taxa including chironomid larvae, enchytraeids, microcrustaceans and nematodes (Bambaradeniya and Amarasinghe, 2003; Kurihara, 1989; Simpson et al., 1993a, 1993b; Weerakoon and Samarasinghe, 1957). Their abundance is highly dependent on rice cultivation practices and may drop to zero during dry intercrop phases (Al-Shami et al., 2010). However, the classical flooded rice paddy invertebrate community tends to quickly recover after relatively short fallow phases with no standing water. Some species are known to be present only for a short time during the flooded phase of rice cultivation. These are often aquatic taxa like chironomid larvae which also include detritivore species (Settle et al., 1996; Stevens et al., 2006). Often, it is hard to make a clear separation between the soil-dwelling and aquatic decomposer fauna in submerged fields as certain taxa (most of which we did not study, e.g. chironomid larvae, Tubificidae, Gastropoda etc.) may partly inhabit both the top soil and water (Kurihara, 1989). Non-flooded agricultural soils in turn harbor a tremendous diversity of predominantly terrestrial microand mesofauna groups (Wardle et al., 2004) including soil nematodes, springtails, mites and enchytraeids (da Silva Moço et al., 2009; Giller, 1996; Sileshi and Mafongoya, 2006). In our study, soil mesofauna abundance was generally higher during dry upland cropping compared to flooded conditions. As a consequence, soil fauna abundance was very low in the fields with continuous flooding (R-WET) throughout the year.

The soil faunal assemblages of flooded rice cultivation systems are adapted to periodic changes in water availability between cropping and fallow phases (Bambaradeniya and Amarasinghe, 2003). The establishment of crop rotations with alternating flooded and non-flooded crops greatly extends dry periods. However, contrary to our expectations the prolonged time of dry phases did not adversely affect the activity of decomposer invertebrates during the succeeding wet phase. This is consistent with the finding that the faunal contribution to litter mass loss was always highest in flooded paddy fields.

We expected a significantly higher abundance of terrestrial invertebrates in the fields with continuous dry cropping (M-DRY); however, we recorded similar shifts in faunal assemblages and abundance between dry and wet seasons in soil of these fields as in those subjected to the rotated moisture regime (M-MIX, R-MIX). This can presumably be attributed to a generally high content of soil moisture during the wet season independent of irrigation or drainage. Hence, soil water content seems to be the major determinant for soil invertebrate abundance and assemblage composition rather than the crop rotations per se. This conclusion is supported by the observation that season (wet or dry) explained most of the variability in the taxonomic composition of soil fauna. Furthermore, the abundance of selected decomposer groups (mites and springtails) was directly or indirectly promoted by lower soil water content in the dry season. Similar effects of climatic seasonality on soil mesofauna abundance have been reported for other tropical agricultural (Badejo, 1990; Singh and Ray, 2015) and forest ecosystems (Boinski and Fowler, 1989). We expected terrestrial taxa to functionally compensate for the decrease of rice straw decomposition by (semi-) aquatic invertebrates during dry cropping. Despite a significantly higher abundance of soil mesofauna in drained fields, their contribution to litter mass loss was negligible. This lack of relationship between faunal assemblages and invertebrate-driven decomposition rates in any season refutes our third hypothesis. Obviously, the highly enhanced microbial decomposition activity compensated (or maybe even overcompensated) the reduced contribution of decomposer invertebrates during dry phases and might have supported the increase in soil fauna abundance. The comparable total amount of decomposed straw during both seasons promotes the general concept of alternating phases of dry cropping and irrigated rice cultivation for a significant reduction of water consumption and greenhouse gas emission.

5. Conclusions

Our study demonstrated that the conversion from continuously flooded rice cultivation to an alternating cropping regime which involves dry upland crops significantly alters the composition of the soil-dwelling fauna as well as its contribution to decomposition processes. Under flooded soil conditions, the invertebratedriven decomposition of rice straw completely compensated for low microbial decay rates. In contrast, during dry cultivation phases, microbial decomposition rates are significantly accelerated while the contribution of invertebrates is of lower magnitude despite an increase in soil invertebrate abundance. This indicates a complementarity of decomposition pathways in different cropping regimes. We conclude that the introduction of non-flooded upland crops into the rice cycle bears only negligible risks for efficient litter decomposition as a fundamental ecosystem service and is therefore a promising approach for increasing sustainability in rice production systems.

Acknowledgements

This study was conducted within the scope of two international projects: ICON ("Introducing Non-Flooded Crops in Rice-Dominated Landscapes: Impact on Carbon, Nitrogen and Water Cycles"; German Research Foundation (DFG): FOR1701/2) and LEGATO ("Land-use intensity and Ecological Engineering – Assessment Tools for risks and Opportunities in irrigated rice based production systems" – www.legato-project.net; Settele et al., 2015). This project was funded by the German Federal Ministry of Education and Research (BMBF, 01LL0917A, 01LL0917L), within the BMBF-Funding Measure "Sustainable Land Management" (http://nachhaltiges-landmanagement.de/en/).

We are grateful for the generous support of the staff members at IRRI (International Rice Research Institute). In particular, we thank Dr. Reiner Wassmann and Jerico Stefan R. Bigornia as well as Liberty Vertudez and Rowena Dela Rosa of IRRI, and Antje Thondorf (UFZ, Dep. of Community Ecology) for technical support. The authors thank Dr. Konstantin Butenko (Skyrabin Institute, Russian Academy of Sciences) for the help with nematode determination. We thank Prof. Dr. Rainer Joergensen (University of Kassel) for determining the ergosterol content on a contract basis. Climate and weather data for this study were kindly provided by the Climate Unit of IRRI.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j. apsoil.2016.05.014.

References

- Acharya, C.N., 1935. Studies on the anaerobic decomposition of plant materials: III. Comparison of the course of decomposition of rice straw under anaerobic, aerobic and partially aerobic conditions. Biochem. I. 29, 1116–1120.
- Al-Shami, S.A., Salmah, M.R.C., Abu Hassan, A., Azizah, M.N.S., 2010. Temporal distribution of larval Chironomidae (Diptera) in experimental rice fields in Penang, Malaysia. J. Asia-Pac. Entomol. 13, 17–22.
- Amato, M., Ladd, J.N., 1992. Decomposition of 14C-labelled glucose and legume material in soils: properties influencing the accumulation of organic residue C and microbial biomass C. Soil Biol. Biochem. 24, 455–464.
- Anderson, J.M., 1995. Soil organisms as engineers: microsite modulation of macroscale processes. In: Jones, C.G., Lawton, J.H. (Eds.), Linking Species & Ecosystems. Springer, US, pp. 94–106.
- Badejo, M.A., 1990. Seasonal abundance of soil mites (Acarina) in two contrasting environments. Biotropica 22, 382–390.
- Baldy, V., Gobert, V., Guerold, F., Chauvet, E., Lambrigot, D., Charcosset, J.Y., 2007. Leaf litter breakdown budgets in streams of various trophic status: effects of dissolved inorganic nutrients on microorganisms and invertebrates. Freshwater Biol. 52, 1322–1335.

Bambaradeniya, C.N.B., Amarasinghe, F.P., 2003. Biodiversity Associated with the Rice Field Agroecosystem in Asian Countries: A Brief Review., Working Paper 63. International Water Management Institute, Colombo Sri Lanka.

Bergfur, J., Friberg, N., 2012. Trade-offs between fungal and bacterial respiration along gradients in temperature, nutrients and substrata: experiments with stream derived microbial communities. Fungal Ecol. 5, 46–52.

Boinski, S., Fowler, N.L., 1989. Seasonal patterns in a tropical lowland forest. Biotropica 21, 223–233.

Bokhorst, S., Wardle, D.A., 2013. Microclimate within litter bags of different mesh size: implications for the 'arthropod effect' on litter decomposition. Soil Biol. Biochem. 58, 147–152.

- Bollen, W.B., 1953. Mulches and soil conditioners: carbon and nitrogen in farm and forest products. Agric. Food Chain. 1, 379–381.
- Bouman, B.A.M., Peng, S., Castañeda, A.R., Visperas, R.M., 2005. Yield and water use of irrigated tropical aerobic rice systems. Agric. Water Manage. 74, 87–105. Bouman, B.A.M., Humphreys, E., Tuong, T.P., Barker, R., 2007. Rice and water. In:
- Donald, L.S. (Ed.), Advances in Agronomy. Academic Press, pp. 187–237. Bozdogan, H., 1987. Model selection and Akaike's Information Criterion (AIC): The
- general theory and its analytical extensions. Psychometrika 52, 345–370. Bradford, M.A., Tordoff, G.M., Eggers, T., Jones, T.H., Newington, J.E., 2002.
- Microbiota, fauna, and mesh size interactions in litter decomposition. Oikos 99, 317–323.
- Cancela da Fonseca, J.P., Sarkar, S., 1998. Soil microarthropods in two different managed ecological systems (Tripura, India). Appl. Soil Ecol. 9, 105–107.
- Cassman, K.G., Pingali, P.L., 1995. Intensification of irrigated rice systems: learning from the past to meet future challenges. GeoJournal 35, 299–305.
- Christensen, B.T., 1985. Wheat and barley straw decomposition under field conditions: effect of soil type and plant cover on weight loss, nitrogen and potassium content. Soil Biol. Biochem. 17, 691–697.
- Clark, M.D., Gilmour, J.T., 1983. The effect of temperature on decomposition at optimum and saturated soil water contents. Soil Sci. Soc. Am. J. 47, 927–929.
- Cobb, N.A., 1918. Estimating the nema population of soil, with special reference to the sugar-beet and root-gall nemas. Heterodera Schachtii Schmidt and Heterodera Radicicola (Greef) Müller, and with a Description of Tylencholaimus Aequalis n. Sp, Agricultural Technology Circular. Bureau of Plant Industry. U.S. Department of Agriculture, Washington, DC, USA.
- Cogle, A.L., Strong, W.M., Saffigna, P.G., Ladd, J.N., Amato, M., 1987. Wheat straw decomposition in subtropical Australia. II: Effect of straw placement on decomposition and recovery of added 15N-urea. Aust. J. Soil Res. 25, 481–490.
- Coleman, D.C., Crossley, D.A., 1996. Fundamentals of Soil Ecology. Academic Press, New York, USA.
- Djajakirana, G., Joergensen, R.G., Meyer, B., 1996. Ergosterol and microbial biomass relationship in soil. Biol. Fert. Soils 22, 299–304.
- Dormann, C.F., Kühn, I., 2008. Angewandte Statistik für die biologischen Wissenschaften. Helmholtz Zentrum für Umweltforschung-UFZ 2.
- FAOSTAT Database, 2008. Rome: Food and Agriculture Organization of the United Nations.
- Frouz, J., Roubíčková, A., Heděnec, P., Tajovský, K., 2015. Do soil fauna really hasten litter decomposition? A meta-analysis of enclosure studies. Eur. J. Soil Biol. 68, 18–24.
- García-Palacios, P., Maestre, F.T., Kattge, J., Wall, D.H., 2013. Climate and litter quality differently modulate the effects of soil fauna on litter decomposition across biomes. Ecol. Lett. 16, 1045–1053.
- Gießelmann, U.C., Martins, K.G., Brändle, M., Schädler, M., Marques, R., Brandl, R., 2010. Diversity and ecosystem functioning: litter decomposition dynamics in the Atlantic Rainforest of Brazil. Appl. Soil Ecol. 46, 283–290.
- Giller, P.S., 1996. The diversity of soil communities, the 'poor man's tropical rainforest'. Biodivers. Conserv. 5, 135–168.
- Graca, M.A.S., 2001. The role of invertebrates on leaf litter decomposition in streams-a review. Int. Rev. Hydrobiol. 86, 383–393.
- Grace, J.B., 2006. Structural Equation Modeling and Natural Systems. Cambridge University Press, New York.
- Haefele, S.M., Banayo, N.P.M., Amarante, S.T., Siopongco, J.D.L.C., Mabesa, R.L., 2013. Characteristics and management options for rice-maize systems in the Philippines. Field Crops Res. 144, 52–61.
- Handa, I.T., Aerts, R., Berendse, F., Berg, M.P., Bruder, A., Butenschoen, O., Chauvet, E., Gessner, M.O., Jabiol, J., Makkonen, M., McKie, B.G., Malmqvist, B., Peeters, E.T.H. M., Scheu, S., Schmid, B., van Ruijven, J., Vos, V.C.A., Hattenschwiler, S., 2014. Consequences of biodiversity loss for litter decomposition across biomes. Nature 509, 218–221.
- Heath, G.W., Edwards, C.A., Arnold, M.K., 1964. Some methods for assessing the activity of soil animals in the breakdown of leaves. Pedobiologia 4, 80–87.
- Heinze, S., Raupp, J., Joergensen, R.G., 2010. Effects of fertilizer and spatial heterogeneity in soil pH on microbial biomass indices in a long-term field trial of organic agriculture. Plant and Soil 328, 203–215.
- Huhta, V., Setälä, H., Haimi, J., 1988. Leaching of N and C from birch leaf litter and raw humus with special emphasis on the influence of soil fauna. Soil Biol. Biochem. 20, 875–878.
- Jansson, S.L., 1958. Tracer studies on nitrogen transformations in soil with special attentron to mineralization-immobilization relationships. Ann. R. Agric. Coll. Sweden 24, 101–361.
- John, K., Jauker, F., Marxsen, J., Zaitsev, A.S., Wolters, V., 2015. Earthworm bioturbation stabilizes carbon in non-flooded paddy soil at the risk of increasing methane emissions under wet soil conditions. Soil Biol. Biochem. 91, 127–132.
- Knapp, E.B., Elliott, L.F., Campbell, G.S., 1983. Microbial respiration and growth during the decomposition of wheat straw. Soil Biol. Biochem. 15, 319–323.

- Koegel-Knabner, I., Amelung, W., Cao, Z., Fiedler, S., Frenzel, P., Jahn, R., Kalbitz, K., Kölbl, A., Schloter, M., 2010. Biogeochemistry of paddy soils. Geoderma 157, 1– 14.
- Kurihara, Y., 1989. Ecology of some ricefields in Japan as exemplified by some benthic fauna, with notes on management. Int. Rev. Hydrobiol. Hydrograph. 74, 507–548.
- Lekha, A., Chopra, G., Gupta, S.R., 1989. Role of soil fauna in decomposition of rice and sorghum straw. Proc. Indian Acad. Sci. 98, 275–284.
- Lepš, J., Šmilauer, P., 2003. Multivariate Analysis of Ecological Data Using CANOCO. Cambridge University Press, New York.
- Leyer, I., Wesche, K., 2007. Multivariate Statistik in Der Ökologie Eine Einführung. Springer, Berlin Heidelberg, New York.
- MacFadyen, A., 1961. Improved funnel-Type extractors for soil arthropods. J. Anim. Ecol. 30, 171–184.
- Manzoni, S., Jackson, R.B., Trofymow, J.A., Porporato, A., 2008. The global stoichiometry of litter nitrogen mineralization. Science 321, 684–686.
- Martinez, A., Larranaga, A., Perez, J., Descals, E., Pozo, J., 2014. Temperature affects leaf litter decomposition in low-order forest streams: field and microcosm approaches. FEMS Microbiol. Ecol. 87, 257–267.
- Merckx, R., den Hartog, A., van Veen, J.A., 1985. Turnover of root-derived material and related microbial biomass formation in soils of different texture. Soil Biol. Biochem. 17, 565–569.
- Mueller, N.D., Gerber, J.S., Johnston, M., Ray, D.K., Ramankutty, N., Foley, J.A., 2012. Closing yield gaps through nutrient and water management. Nature 490, 254– 257.
- Oksanen, J., Kindt, R., Legendre, P., O'Hara, B., Stevens, M.H.H., Oksanen, M.J., Suggests, M., 2007. The Vegan Package. Community ecology package.
- Perez, J., Galan, J., Descals, E., Pozo, J., 2014. Effects of fungal inocula and habitat conditions on alder and eucalyptus leaf litter decomposition in streams of northern Spain. Microb. Ecol. 67, 245–255.
- Rao, C.R., 1964. The use and interpretation of principal component analysis in applied research. Sankhyā: Indian J. Stat. Ser. A 26, 329–358.
- Reddy, M.V., Reddy, V.R., Yule, D.F., Cogle, A.L., George, P.J., 1994. Decomposition of straw in relation to tillage, moisture, and arthropod abundance in a semi-arid tropical Alfisol. Biol. Fert. Soils 17, 45–50.
- Saha, P.K., Miah, M.A.M., Hossain, A.T.M.S., Rahman, F., Saleque, M.A., 2009. Contribution of rice straw to potassium supply in rice-fallow-rice cropping pattern. Bangladesh J. Agric. Res. 34, 633–643.
- Santos, P.F., Whitford, W.G., 1981. The effects of microarthropods on litter decomposition in a chihuahuan desert ecosystem. Ecology 62, 654–663.
- Schädler, M., Brandl, R., 2005. Do invertebrate decomposers affect the disappearance rate of litter mixtures? Soil Biol. Biochem. 37, 329–337.
- Schädler, M., Jung, G., Auge, H., Brandl, R., 2003. Palatability, decomposition and insect herbivory: patterns in a successional old-field plant community. Oikos 103, 121–132.
- Schmidt, A., Auge, H., Brandl, R., Heong, K.L., Hotes, S., Settele, J., Villareal, S., Schädler, M., 2015a. Small-scale variability in the contribution of invertebrates to litter decomposition in tropical rice fields. Basic Appl. Ecol. 16, 674–680.
- Schmidt, A., John, K., Arida, G., Auge, H., Brandl, R., Horgan, F.G., Hotes, S., Marques, L., Radermacher, N., Settele, J., Wolters, V., Schädler, M., 2015b. Effects of residue management on decomposition in irrigated rice fields are not related to changes in the decomposer community. PLoS One 10 (7), e0134402.
- Setälä, H., Haimi, J., Huhta, V., 1988. A microcosm study on the respiration and weight loss in birch litter and raw humus as influenced by soil fauna. Biol. Fert. Soils 5, 282–287.
- Settele, J., Spangenberg, J.H., Heong, K.L., Burkhard, B., Bustamante, J.V., Cabbigat, J., Van Chien, H., Escalada, M., Grescho, V., Hai, L.H., Harpke, A., Horgan, F.G., Hotes, S., Jahn, R., Kühn, I., Marquez, L., Schädler, M., Tekken, V., Vetterlein, D., Villareal, S.B., Westphal, C., Wiemers, M., 2015. Agricultural landscapes and ecosystem services in South-East Asia—the LEGATO-Project. Basic Appl. Ecol. 16, 661–664. Settle, W.H., Ariawan, H., Astuti, E.T., Cahyana, W., Hakim, A.L., Hindayana, D.,
- Settle, W.H., Ariawan, H., Astuti, E.T., Cahyana, W., Hakim, A.L., Hindayana, D., Lestari, A.S., 1996. Managing tropical rice pests through conservation of generalist natural enemies and alternative prey. Ecology 77 (7), 1975–1988.
- Sileshi, G., Mafongoya, P.L., 2006. Variation in macrofaunal communities under contrasting land use systems in eastern Zambia. Appl. Soil Ecol. 33, 49–60.
- Simpson, I.C., Roger, P.A., Oficial, R., Grant, I.F., 1993a. Density and composition of aquatic oligochaete populations in different farmers' ricefields. Biol. Fert. Soils 16, 34–40.
- Simpson, I.C., Roger, P.A., Oficial, R., Grant, I.F., 1993b. Impacts of agricultural practices on aquatic oligochaete populations in ricefields. Biol. Fert. Soils 16, 27– 33.
- Singh, L.A., Ray, D.C., 2015. Effect of no-tillage and tillage on the ecology of mite, Acarina (Oribatida) in two different farming systems of paddy field in Cachar district of Assam. J. Environ. Biol. 36, 319–323.
- Singh, Y., Singh, B., Timsina, J., 2005. Crop residue management for nutrient cycling and improving soil productivity in rice-based cropping systems in the tropics. Adv. Agron. 85, 269–407.
- Singh, Y., Gupta, R.K., Singh, J., Singh, G., Singh, G., Ladha, J.K., 2010. Placement effects on rice residue decomposition and nutrient dynamics on two soil types during wheat cropping in rice-wheat system in northwestern India. Nutr. Cycl. Agroecosyst. 88, 471–480.
- Steinberger, Y., Whitford, W.G., 1988. Decomposition process in Negev ecosystems. Oecologia 75, 61–66.
- Stevens, M.M., Helliwell, S., Cranston, P.S., 2006. Larval chironomid communities (Diptera: chironomidae) associated with establishing rice crops in Southern New South Wales, Australia. Hydrobiologia 556, 317–325.

Talbot, J.M., Treseder, K.K., 2011. Dishing the dirt on carbon cycling. Nature Clim. Change 1, 144–146.

- Tian, G., Kang, B.T., Brussaard, L., 1992. Biological effects of plant residues with contrasting chemical compositions under humid tropical conditions – decomposition and nutrient release. Soil Biol. Biochem. 24, 1051–1060.
- Tian, G., Brussaard, L., Kang, B.T., 1993. Biological effects of plant residues with contrasting chemical compositions under humid tropical conditions: effects on soil fauna. Soil Biol. Biochem. 25, 731–737.
- Tian, G., Brussaard, L., Kang, B.T., 1995. An index for assessing the quality of plant residues and evaluating their effects on soil and crop in the (sub-)humid tropics. Appl. Soil Ecol. 2, 25–32.
- Timsina, J., Jat, M., Majumdar, K., 2010. Rice-maize systems of South Asia: current status, future prospects and research priorities for nutrient management. Plant and Soil 335, 65–82.
- Timsina, J., Buresh, R.J., Dobermann, A., Dixon, J., 2011. Rice-maize Systems in Asia: Current Situation and Potential. International Rice Research Institute (IRRI) International Maize and Wheat Improvement Center, Los Baños, Philippines. USDA, 2007. US Department of Agriculture. www.usda.gov.
- Vigil, M.F., Kissel, D.E., 1991. Equations for estimating the amount of nitrogen mineralized from crop residues. Soil Sci. Soc. Am. J. 55, 757–761.
- Wall, D.H., Bradford, M.A., St. John, M.G., Trofymow, J.A., Behan-Pelletier, V., Bignell, D.E., Dangerfield, J.M., Parton, W.J., Rusek, J., Voigt, W., Wolters, V., Gardel, H.Z., Ayuke, F.O., Bashford, R., Beljakova, O.I., Bohlen, P.J., Brauman, A., Flemming, S., Henschel, J.R., Johnson, D.L., Jones, T.H., Kovarova, M., Kranabetter, J.M., Kutny, L. E.S., Lin, K.-C., Maryati, M., Masse, D., Pokarzhevskii, A., Rahman, H., Sabar, M.G., Salamon, J.-A., Swift, M.J., Varela, A., Vasconcelos, H.L., White, D.O.N., Zou, X., 2008. Global decomposition experiment shows soil animal impacts on decomposition are climate-dependent. Global Change Biol. 14, 2661–2677. doi: http://dx.doi.org/10.1111/j.1365-2486.2008.01672.x.
- Wardle, D.A., Nicholson, K.S., Bonner, K.I., Yeates, G.W., 1999. Effects of agricultural intensification on soil-associated arthropod population dynamics, community

structure, diversity and temporal variability over a seven-year period. Soil Biol. Biochem. 31, 1691–1706.

- Wardle, D.A., Bardgett, R.D., Klironomos, J.N., Setälä, H., van der Putten, W.H., Wall, D.H., 2004. Ecological linkages between aboveground and belowground biota. Science 304, 1629–1633.
- Wassmann, R., Vlek, P.L.G., 2004. Mitigating greenhouse gas emissions from tropical agriculture: scope and research priorities. environment. Dev. Sustain. 6, 1–9.
- Watanabe, I., Roger, P.-A., 1985. Ecology of flooded rice fields, Wetland Soils: Characterization, Classification, and Utilization, Los Baños, Philippines, pp. 229– 243.
- Weerakoon, A.C.J., Samarasinghe, E.L., 1957. Mesofauna of the soil of a paddy-field in Ceylon – a preliminary survey. Ceylon J. Sci. (Biological Science) 1, 155–170.
- Widyastuti, R., 2002. Soil fauna in rainfed paddy field ecosystems: their role in organic matter decomposition and nitrogen mineralization. Ecology and Development Series Band 3. Cuvillier Verlag, Göttingen.
- Witt, C., Cassman, K.G., Olk, D.C., Biker, U., Liboon, S.P., Samson, M.I., Ottow, J.C.G., 2000. Crop rotation and residue management effects on carbon sequestration, nitrogen cycling and productivity of irrigated rice systems. Plant and Soil 225, 263–278.
- Witt, C., Pasuquin, J.M.C.A., Mutters, R., Buresh, R.J., 2005. New leaf color chart for effective nitrogen management in rice. Better crops 89, 36–39.
- Wolters, V., 1991. Soil invertebrates effects on nutrient turnover and soil structure – a review. Zeitschrift für Pflanzenernährung und Bodenkunde 154, 389–402.
- Yeates, G.W., Bongers, T., De Goede, R.G.M., Freckman, D.W., Georgieva, S.S., 1993. Feeding habits in soil nematode families and genera – an outline for soil ecologists. J. Nematol. 25, 315.
- da Silva Moço, M.K., da Gama-Rodrigues, E.F., da Gama-Rodrigues, A.C., Machado, R. C.R., Baligar, V.C., 2009. Soil and litter fauna of cacao agroforestry systems in Bahia, Brazil. Agroforest Syst. 76, 127–138.
- van den Wollenberg, A.L., 1977. Redundancy analysis an alternative for canonical correlation analysis. Psychometrika 42, 207–219.